

TAXONOMY AND IDENTIFICATION OF *CANDIDA* AND *CRYPTOCOCCUS*. CONVENTIONAL METHOD AND AUTOMATION SYSTEMS

長庚醫院 醫檢部

郭安靜

2016/08/27

OUTLINE

- General characteristics
- Yeast identification
- *Candida*
- *Cryptococcus*
- Other yeast
 - *Trichosporon, Rhodotorula*
 - *Malassezia, Prototheca*

DEFINITION OF YEASTS

- Eukaryote –cells have defined nucleus and nuclear membrane
- Single cell (vs mould mostly multicellular and forms mycelium) that reproduces by budding
- Reproduce by **sexual** and **asexual** process
- Yeast- appearance is generally moist looking with smooth round colonies

IDENTIFICATION OF YEAST

○ Conventional method

- Gross morphology
- Microscopic morphology (wet mount, Gram stain, India ink)
- Germ tube
- Morphology, cornmeal tween 80 agar (Dalmau plate)
- **Chromogenic agar**
- Biochemical tests (Assimilation, API/ATB, Vitek etc, Urease, Phenol oxidase)

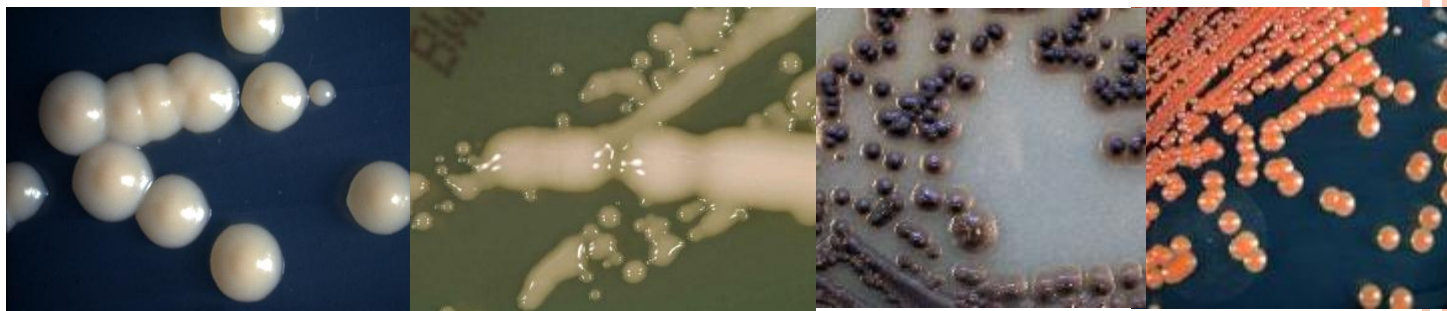
○ DNA sequencing

○ **MALDI-TOF mass spectrometry**

GROSS MORPHOLOGY

Table 8.5–1 General list of yeast species or genera based on colony color^a

White, cream, or tan	Brown or black	Salmon, pink, or red
<i>Blastoschizomyces</i>	<i>Aureobasidium</i>	<i>Rhodotorula</i>
<i>Candida</i>	<i>Phaeoannellomyces elegans</i>	<i>Sporobolomyces</i>
<i>Cryptococcus</i>	<i>Hortaea werneckii</i> ^b	
<i>Geotrichum</i> ^d	<i>Phaeococcomyces exophialae</i> ^c	
<i>Hansenula</i>	<i>Ustilago</i>	
<i>Kloeckera</i>		
<i>Malassezia</i>		
<i>Prototheca</i> ^d		
<i>Saccharomyces</i>		
<i>Trichosporon</i>		
<i>Ustilago</i>		
Teleomorphs ^c		
<i>Debaryomyces</i>		
<i>Kluyveromyces</i>		
<i>Pichia</i>		



^a Based on growth on Sabouraud glucose agar (Emmon's modification), potato dextrose agar, malt extract agar, or BHI agar. Note that all *Malassezia* spp. except *Malassezia pachydermatis* require exogenously supplied lipid (e.g., olive oil) for growth.

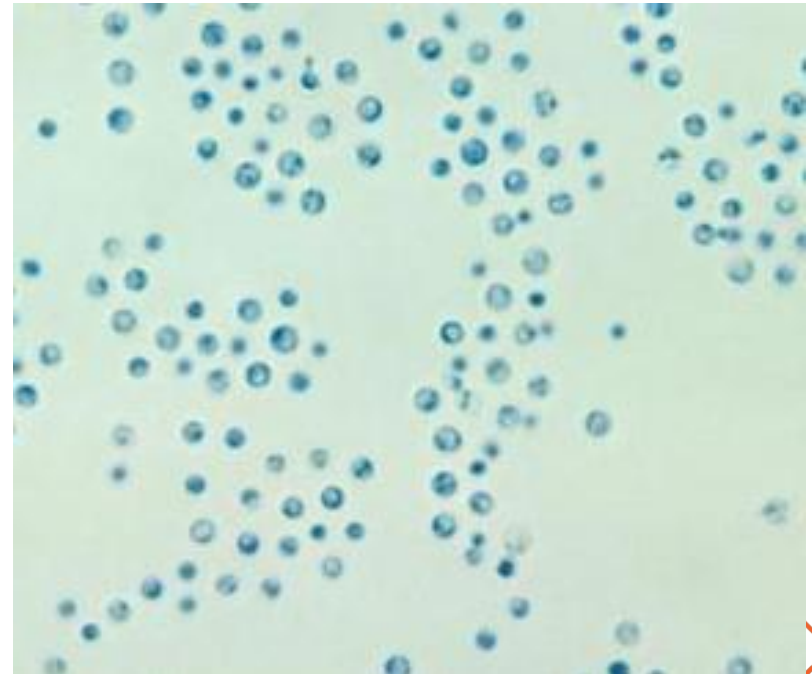
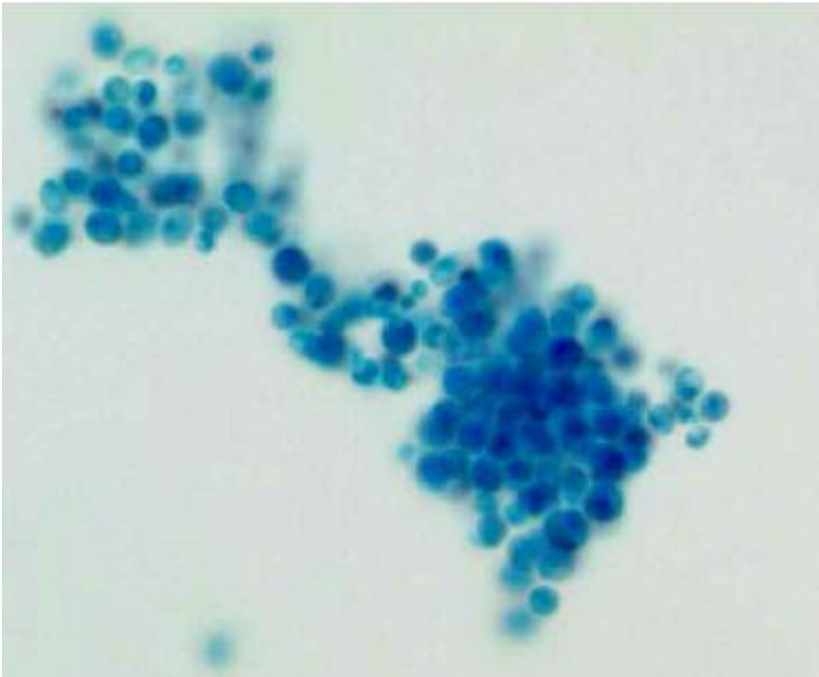
^b Previously *Exophiala werneckii*.

^c A second anamorphic (asexual) form (i.e., synanamorph) of *Exophiala (Wangiella) dermatitidis*.

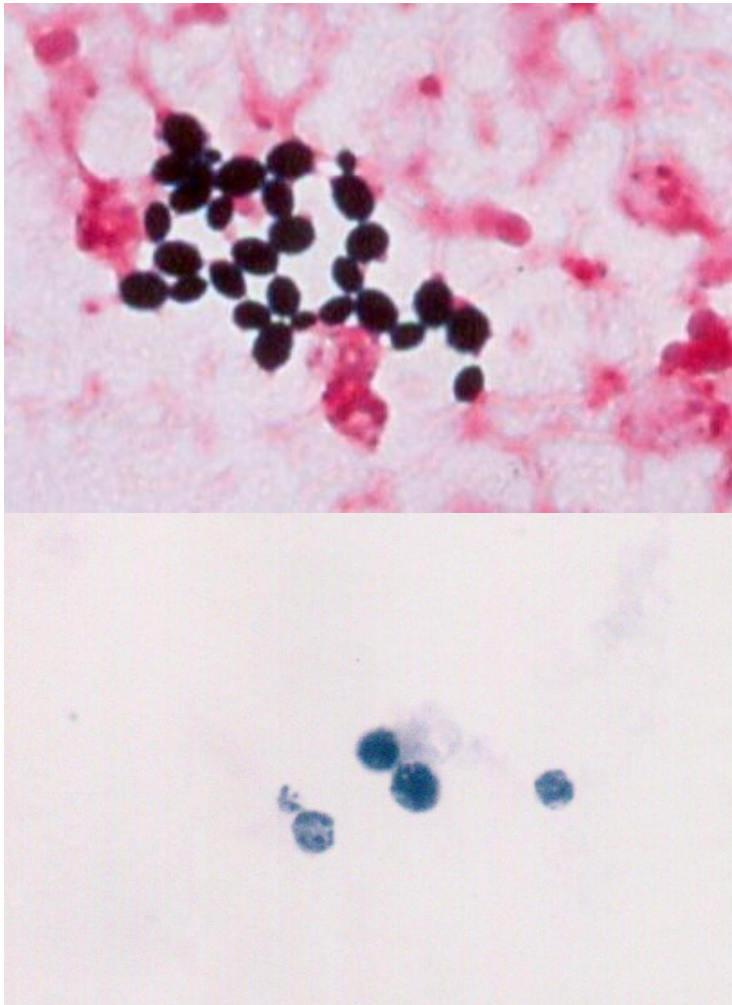
^d Not a yeast but produces yeastlike colonies on mycological media.

MICROSCOPIC EXAMINATION

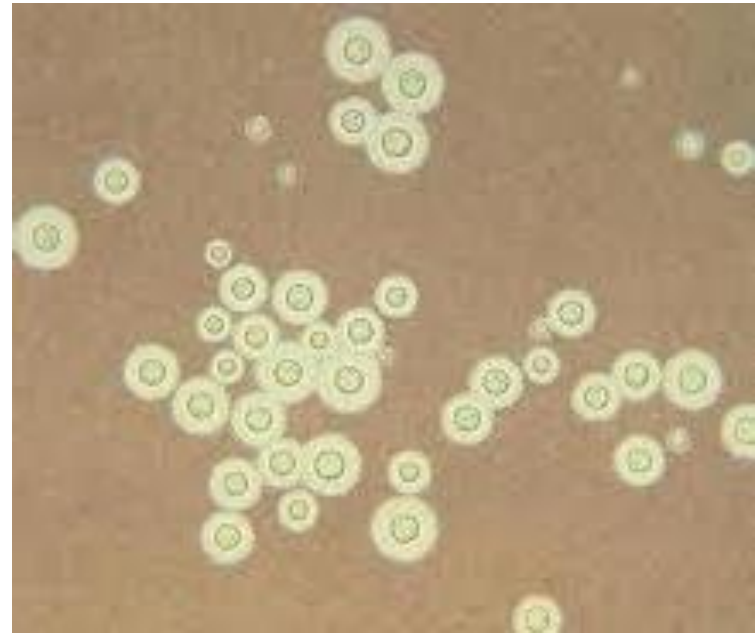
- Do microscopic examination -perform a wet mount or Gram stain
 - Methylene blue, lactophenocotton blue



GRAM STAIN

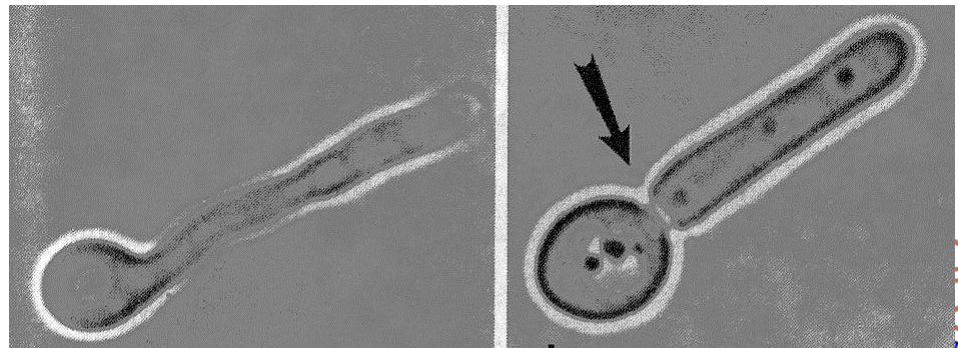


INDIAN INK



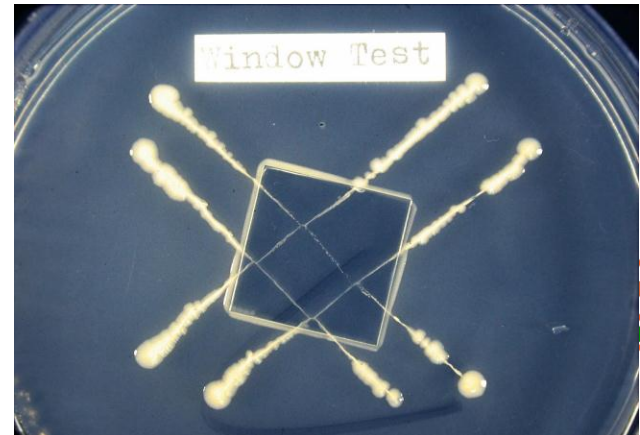
GERM TUBE TEST

- Procedure –inoculate a light inoculum of yeast from a young culture into 0.5 mL serum, incubate at 35°C for 2-3 hours. Look for tube like structures appearing from blastoconidia (no constriction at the point of formation).
- *Candida albicans* and *C. dubliniensis* form **germ tubes**.



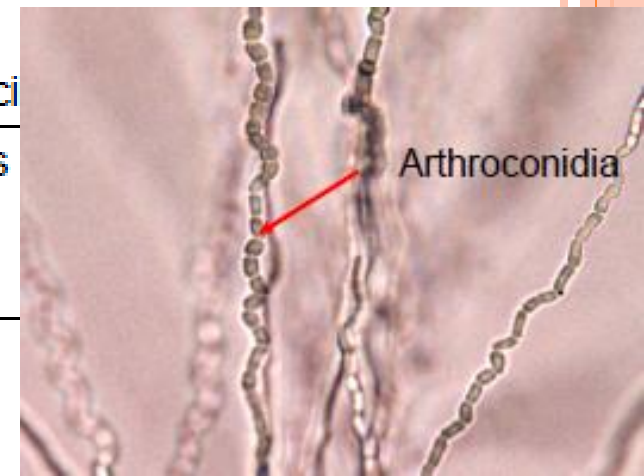
YEAST MORPHOLOGY

- Perform yeast morphology. Use **Cornmeal agar with 1% Tween 80** (Dalmau plate).
 - **chlamydospore** (chlamydoconidia) –tentatively identifies as *Candida albicans*.
 - Other features seen include **blastoconidia**, **pseudohyphae**, **hyphae**, **arthroconidia**.
 - Incubate at 22-26°C for 24 hours, or 48 hours if necessary.
- Can narrow down to genus level.



YEAST MORPHOLOGY

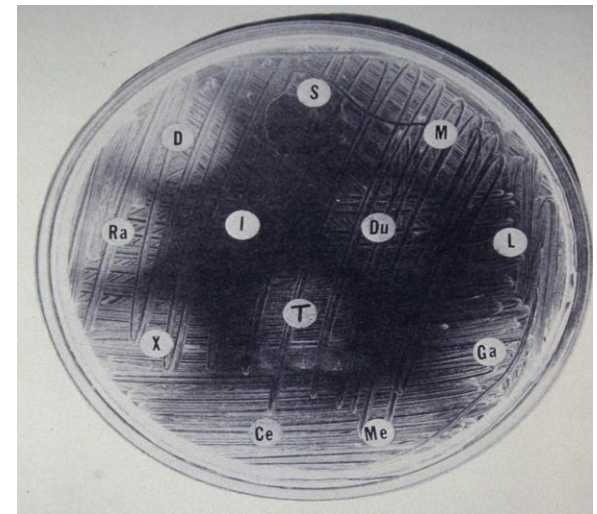
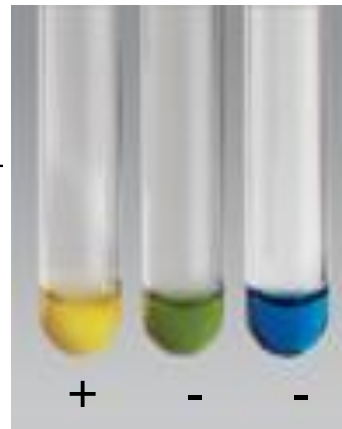
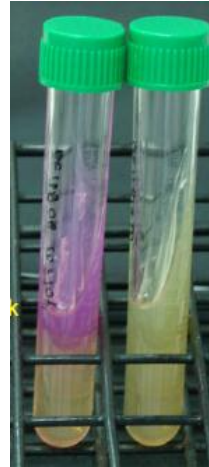
Structures observed	Probable Identification
Pseudohyphae (occasional hyphae) Blastoconidia Chlamydospores	Candida albicans Candida dubliniensis
Pseudohyphae (occasional hyphae) Blastoconidia	Candida species
Blastoconidia	Candida species (C. glabrata) Cryptococcus species Rhodotorula species Saccharomyces species
Hyphae & pseudohyphae Blastoconidia Arthroconidia	Trichosporon species
Hyphae Arthroconidia	Geotrichum species



BIOCHEMICAL TEST

○ Biochemical tests

- Urease,
- Phenol oxidase
 - Caffeic acid agar/Bird seed agar/niger seed agar
- Rapid trehalose
- Carbon assimilation
- API/ATB
- Vitek 2



CHROMOGENIC AGAR

- If resources or expertise are limited.
- mixed culture with more than 1 type of *Candida* spp.

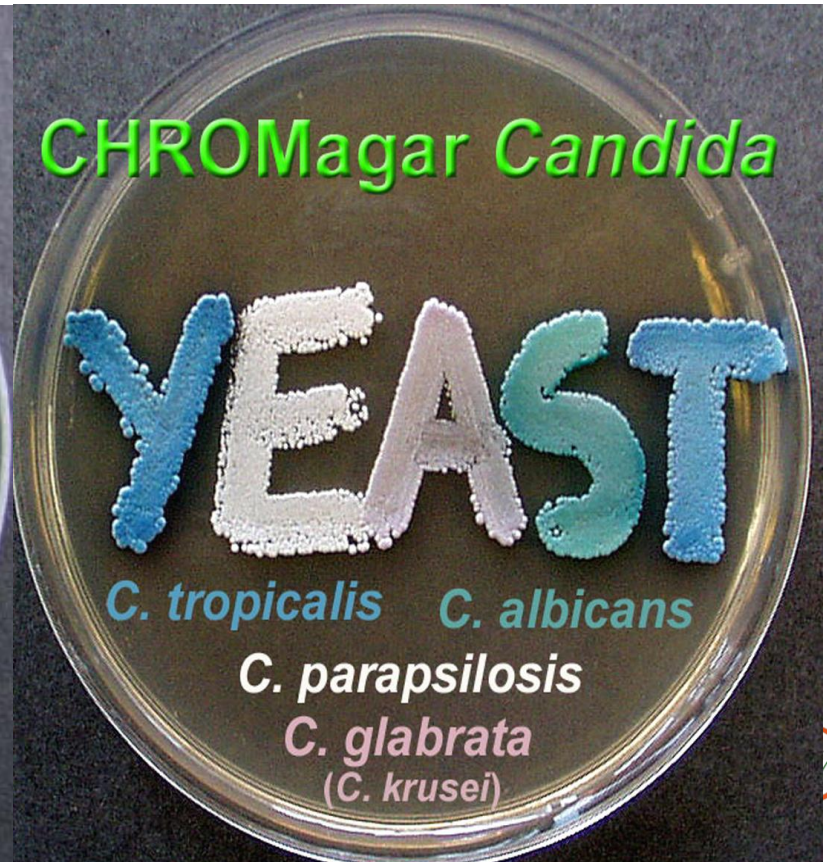
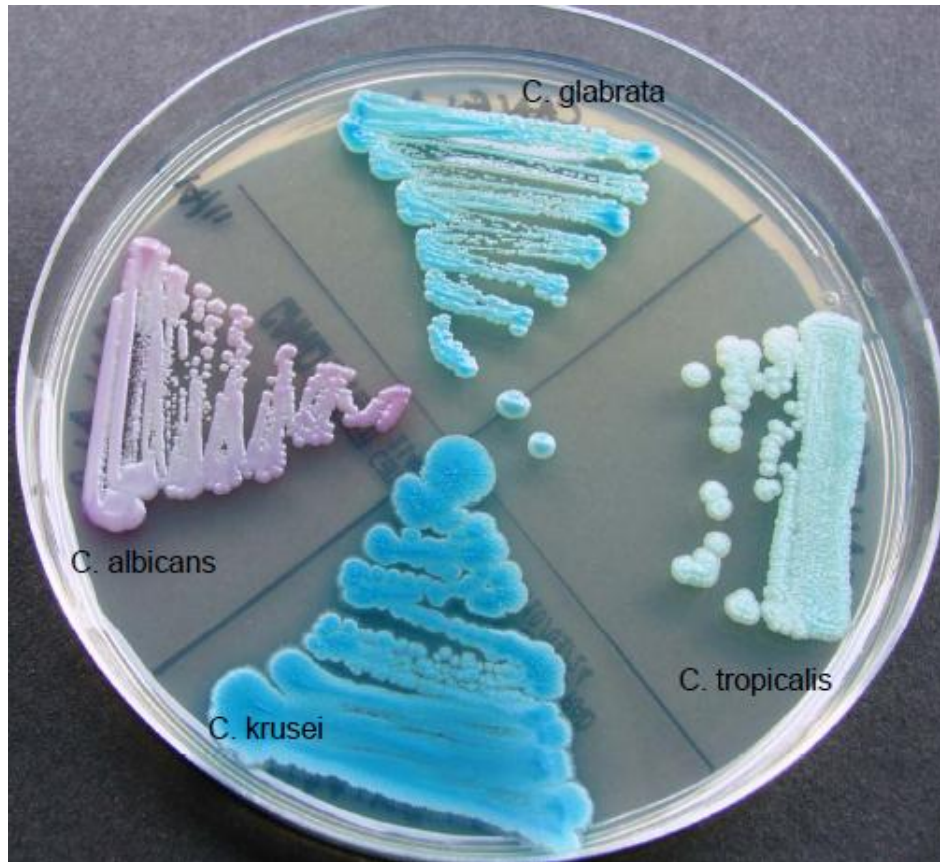


Table 8.6–1 Presumptive identification tests for yeasts on primary culture

Test	Item no. ^a	Organism
CHROMagar	8.5.VI.A	<i>C. krusei</i> , <i>C. albicans</i> , <i>C. tropicalis</i> , <i>Trichosporon</i> spp.
Germ tube	8.6.IV.A	<i>C. albicans</i> complex ^b
<i>C. albicans</i> screen	8.6.IV.B	<i>C. albicans</i> , <i>C. dubliniensis</i>
Rapid urease	8.6.IV.C	<i>Cryptococcus</i> spp., <i>Rhodotorula</i> spp., <i>Trichosporon</i> spp. (variable), <i>C. krusei</i> (variable), <i>Malassezia pachydermatis</i> , <i>Candida lipolytica</i> (i.e., basidiomycetous yeasts)
Rapid nitrate reductase	8.6.IV.D	<i>C. albidus</i> (+), <i>C. neoformans</i> (–), <i>C. terreus</i> (+)
Caffeic acid disk	8.6.IV.E	<i>C. neoformans</i>
Rapid trehalose assimilation	8.6.IV.F	<i>C. glabrata</i>
India ink	8.3.V.D	<i>Cryptococcus</i> spp.

^a In this handbook.

^b This complex consists of *C. albicans*, *C. dubliniensis*, *C. stellatoidea* (obsolete), and *Candida africanum* sp. nom. inval.

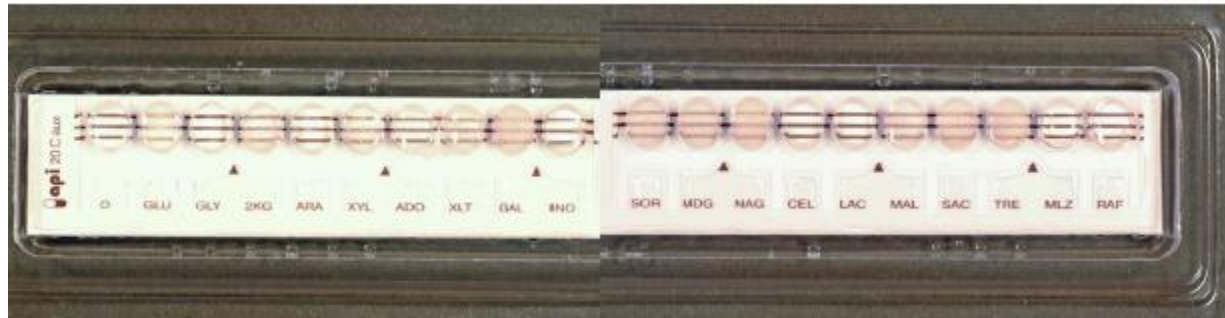
CARBON ASSIMILATION

Table 8.8–4 Culture and biochemical characteristics of yeasts frequently isolated from clinical specimens^a

Species	Growth at 37°C	Pellicle in broth	Pseudo- or true hyphae	Chlamydospores	Germ tubes	Capsule, India ink	Assimilation of ^b :											Fermentation of:						Urease	KNO ₃	Phenol oxidation	Ascospores	
							Glucose	Maltose	Sucrose	Lactose	Galactose	Melibiose	Cellobiose	Inositol	Xylose	Raffinose	Trehalose	Dulcitol	Glucose	Maltose	Sucrose	Lactose	Galactose					Trehalose
<i>C. albicans</i>	+	—	+	+	+	—	+	+	+	—	+	—	—	+	—	+	—	F	F	—	—	F	F	—	—	—	—	—
<i>Candida catenulata</i>	+	—	+	—	—	—	+	+	—	—	+	—	—	+	—	—	—	F	—	—	—	—	—	—	—	—	—	—
<i>C. dubliniensis</i>	+	—	+	+	+	—	+	+	+	—	+	—	—	+	—	+	—	F	F	—	—	F	F	—	—	—	—	—
<i>Candida famata</i>	+	—	—	—	—	—	+	+	+	+	+	+	—	+	+	+	+	W	—	W	—	—	W	—	—	—	+	
<i>C. glabrata</i>	+	—	—	—	—	—	+	+	—	—	—	—	—	—	—	+	—	F	—	—	—	—	F	—	—	—	—	
<i>Candida guilliermondii</i>	+	—	+	—	—	—	+	+	+	—	+	+	—	+	+	+	+	F	—	F	—	F	F	—	—	—	—	+
<i>Candida inconspicua</i>	+	—	—	—	—	—	+	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—	—
<i>C. kefyr</i>	+	—	+	—	—	—	+	—	+	+	—	+	—	+	+	—	—	F	—	F	F	F	—	—	—	—	—	—
<i>Candida krusei</i> ^d	+	+	+	—	—	—	+	—	—	—	—	—	—	—	—	—	—	F	—	—	—	—	—	+	—	—	—	+
<i>Candida lambica</i>	+	+	+	—	—	—	+	—	—	—	—	—	—	+	—	—	—	F	—	—	—	—	—	—	—	—	—	+



API/ATB-API 20C AUX /ATB 32C



ID 32 C V3.0

<i>Candida albicans 1</i>
<i>Candida albicans 2</i>
<i>Candida boidinii</i>
<i>Candida catenulata</i>
<i>Candida colliculosa</i>
<i>Candida dattila</i>
<i>Candida dubliniensis</i>
<i>Candida famata</i>
<i>Candida glabrata</i>
<i>Candida globosa</i>
<i>Candida guilliermondii</i>
<i>Candida hellenica</i>
<i>Candida holmii</i>
<i>Candida inconspicua/norvegensis</i>
<i>Candida intermedia</i>
<i>Candida kefyr</i>
<i>Candida krusei</i>
<i>Candida lambica</i>
<i>Candida lipolytica</i>
<i>Candida lusitanae</i>
<i>Candida magnoliae</i>
<i>Candida melibiosica</i>
<i>Candida membranifaciens</i>
<i>Candida norvegica</i>
<i>Candida parapsilosis</i>
<i>Candida pelliculosa</i>
<i>Candida pulcherrima</i>
<i>Candida rugosa</i>
<i>Candida sake</i>

<i>Candida silvicola</i>
<i>Candida sphaerica</i>
<i>Candida tropicalis</i>
<i>Candida utilis</i>
<i>Candida valida</i>
<i>Candida zeylanoides</i>
<i>Cryptococcus albidus</i>
<i>Cryptococcus curvatus</i>
<i>Cryptococcus humicola</i>
<i>Cryptococcus laurentii</i>
<i>Cryptococcus neoformans</i>
<i>Cryptococcus terreus</i>
<i>Cryptococcus uniguttulatus</i>
<i>Debaryomyces etchellsii/carsonii</i>
<i>Debaryomyces polymorphus</i>
<i>Geotrichum capitatum</i>
<i>Geotrichum spp</i>
<i>Kloeckera apis/apiculata</i>
<i>Kloeckera japonica</i>
<i>Kodamaea ohmeri</i>
<i>Pichia farinosa</i>
<i>Rhodotorula glutinis</i>
<i>Rhodotorula minuta</i>
<i>Rhodotorula mucilaginosa</i>
<i>Saccharomyces cerevisiae</i>
<i>Saccharomyces kluyverii</i>
<i>Sporobolomyces salmonicolor</i>
<i>Stephanoascus ciferrii</i>
<i>Trichosporon inkin</i>
<i>Trichosporon asahii</i>
<i>Trichosporon mucoides</i>
<i>Williopsis saturnus</i>
<i>Zygosaccharomyces spp</i>

API 20 C AUX V4.0

<i>Candida albicans 1</i>
<i>Candida albicans 2</i>
<i>Candida boidinii</i>
<i>Candida colliculosa</i>
<i>Candida dubliniensis</i>
<i>Candida famata</i>
<i>Candida glabrata</i>
<i>Candida guilliermondii</i>
<i>Candida kefyr</i>
<i>Candida krusei/inconspicua</i>
<i>Candida lusitanae</i>
<i>Candida magnoliae</i>
<i>Candida norvegensis</i>
<i>Candida parapsilosis</i>
<i>Candida pelliculosa</i>
<i>Candida rugosa</i>
<i>Candida sphaerica 1</i>
<i>Candida sphaerica 2</i>
<i>Candida tropicalis</i>
<i>Candida utilis</i>
<i>Candida zeylanoides</i>

<i>Cryptococcus albidus</i>
<i>Cryptococcus humicola</i>
<i>Cryptococcus laurentii</i>
<i>Cryptococcus neoformans</i>
<i>Cryptococcus terreus</i>
<i>Cryptococcus uniguttulatus</i>
<i>Geotrichum capitatum</i>
<i>Geotrichum klebahnii</i>
<i>Kloeckera spp</i>
<i>Kodamaea ohmeri</i>
<i>Pichia angusta</i>
<i>Prototheca wickerhamii</i>
<i>Rhodotorula glutinis</i>
<i>Rhodotorula minuta</i>
<i>Rhodotorula mucilaginosa 1</i>
<i>Rhodotorula mucilaginosa 2</i>
<i>Saccharomyces cerevisiae 1</i>
<i>Saccharomyces cerevisiae 2</i>
<i>Sporobolomyces salmonicolor</i>
<i>Stephanoascus ciferrii</i>
<i>Trichosporon asahii</i>
<i>Trichosporon inkin</i>
<i>Trichosporon mucoides</i>

Malassezia ??

VITEK



Disadvantage -not good for some yeasts
eg *Trichosporon* misidentified as *Cryptococcus laurentii*

Vitek® 2 YST ID Card

YEAST

Candida albicans

Candida boidinii

Candida catenulata

Candida colliculosa

Candida dubliniensis

Candida famata

Candida freyschussii

Candida glabrata

Candida guilliermondii

Candida haemulonii

Candida inconspicua

Candida intermedia

Candida kefyr

Candida krusei

Candida lambica

Candida lipolytica

Candida lusitanae

Candida magnoliae

Candida norvegensis

Candida parapsilosis

Candida pelliculosa

Candida pulcherrima

Candida rugosa

Candida sake

Candida sphaerica

Candida tropicalis

Candida utilis

Candida zeylanoides

Cryptococcus albidus

Cryptococcus laurentii

Cryptococcus neoformans

Cryptococcus terreus

Cryptococcus uniguttulatus

Geotrichum capitatum

Geotrichum klebahnii

Kloeckera spp

Malassezia furfur

Malassezia pachydermatis

Pichia farinosa

Kodamaea ohmeri

Prototheca wickerhamii

Prototheca zopfii

Rhodotorula glutinis

Rhodotorula minuta

Rhodotorula mucilaginosa

Saccharomyces cerevisiae

Sporobolomyces salmonicolor

Stephanoascus ciferrii

Trichosporon asahii

Trichosporon inkin

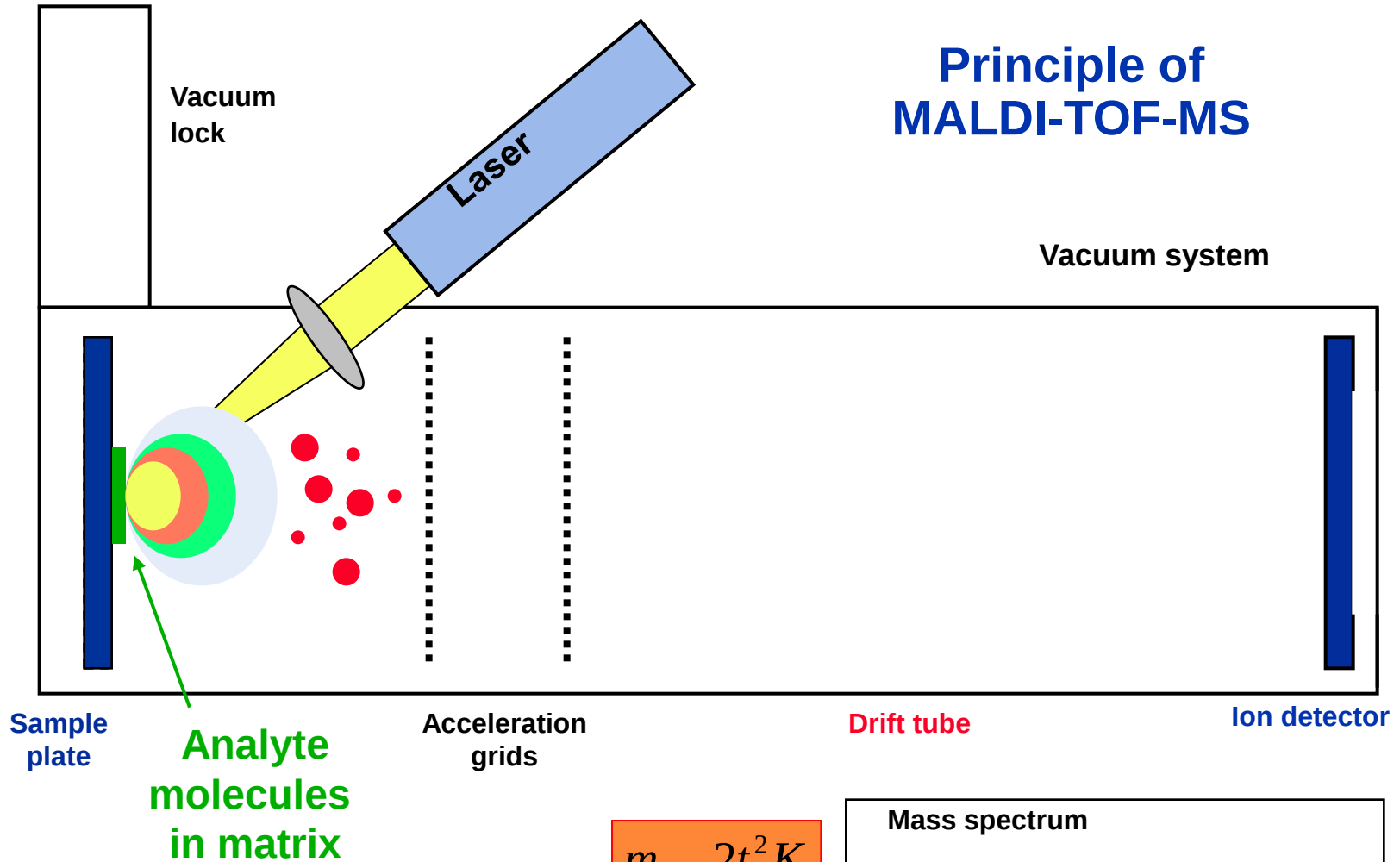
Trichosporon mucoides

Zygosaccharomyces bailii

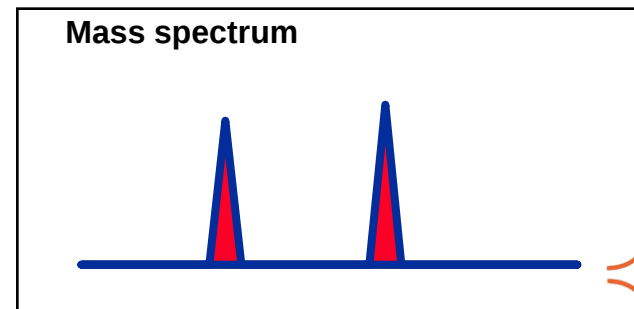
MALDI-TOF MS

- Matrix-assisted laser desorption/ionization time of flight mass spectrometry
- Used to analyze samples of many types, including solutions of organic molecules, nucleic acids, proteins, and whole microorganisms

Principle of MALDI-TOF-MS



$$\frac{m}{z} = \frac{2t^2K}{L^2}$$



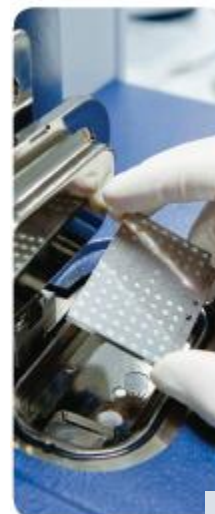
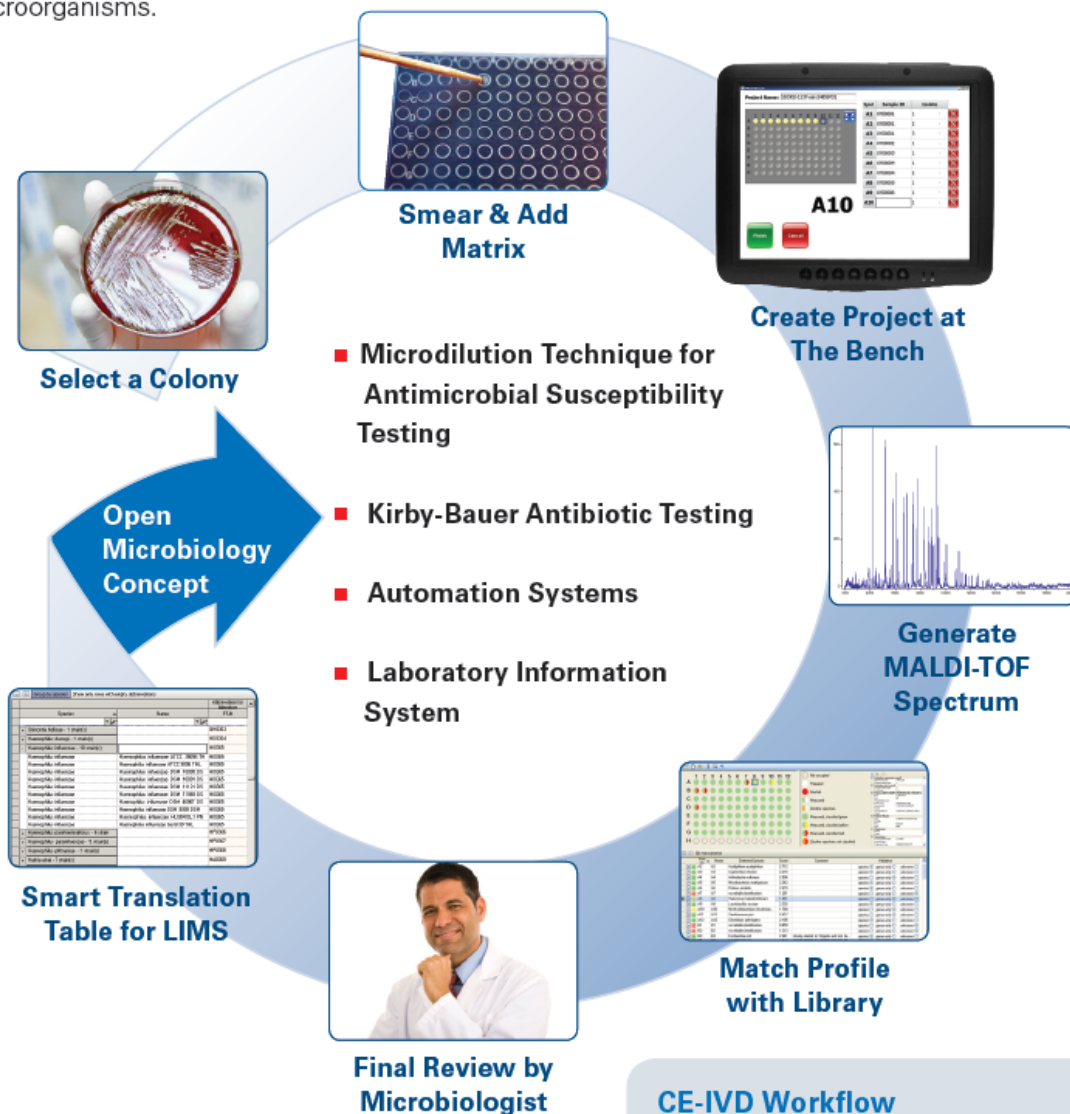
MALDI-TOF MS

- **MALDI-Biotyper system**- Bruker Daltonics (Leipzig, Germany) (who have recently partnered with Becton-Dickinson, Franklin Lakes, NJ)
- **Vitex-MS** - Shimadzu Corp. (Kyoto, Japan) and bioMerieux SA (Marcyl'Etoile, France)



MALDI-BIOTYPER SYSTEM

microorganisms.



MALDI-TOF database

- *Candida* spp. - >60
- *Cryptococcus* spp. – 20
- *Geotricum* spp. -3
- *Saccharomyces*
- *Malassezia* spp
- *Pichia* spp. – 10
- *Prototheca*
- *Rhodotorula* spp – 6
- *Trichosporon* spp -20
- *Alternaria alternata*
- *Aspergillus* spp. – 8
- *Epidermophyton*
- *Exophiala*
- *Fusarium* -2
- *Microsporum* spp. –2
- *Mucor*
- *Paecilomyces* - 2
- *Penicillium* spp. – 3
- *Phoma* - 2
- *Rhizopus*
- *Sporothrix*
- *Trichophyton* spp - 4

YEAST + formic acid

Meaning of Score Values

Range	Description	Yeast
2.300 ... 3.000	highly probable species identification	81
2.000 ... 2.299	secure genus identification, probable species identification	77.8%
1.700 ... 1.999	probable genus identification	16.0%
0.000 ... 1.699	not reliable identification	6.2%

Table 2 Routine identification performance of matrix-assisted laser desorption/ionization time-of-flight mass spectrometry for yeasts and nontuberculous mycobacteria (NTM).

Microorganism	Total no.	No. (%) of isolates identified to		No. (%) of unidentified isolates		
		Species level	Genus level	Total	No peaks found	No reliable identification
Yeast	81	63 (77.8)	76 (93.8)	5 (6.2)	4 (4.9)	1 (1.2)
<i>Candida glabrata</i>	19	19 (100.0)	19 (100.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Candida tropicalis</i>	16	12 (75.0)	15 (93.7)	1 (6.3)	1 (6.3)	0 (0.0)
<i>Candida albicans</i>	15	11 (73.3)	15 (100.0)	0 (0.0)	0 (0.0)	0 (0.0)
<i>Candida parapsilosis</i>	14	9 (64.3)	12 (85.7)	2 (14.3)	2 (14.3)	0 (0.0)
Others	17	12 (70.6)	15 (88.2)	2 (11.8)	1 (5.9)	1 (5.9)

APPLICATION OF MALDI-TOF MS TO THE IDENTIFICATION OF YEASTS IN THE CLINICAL LABORATORY

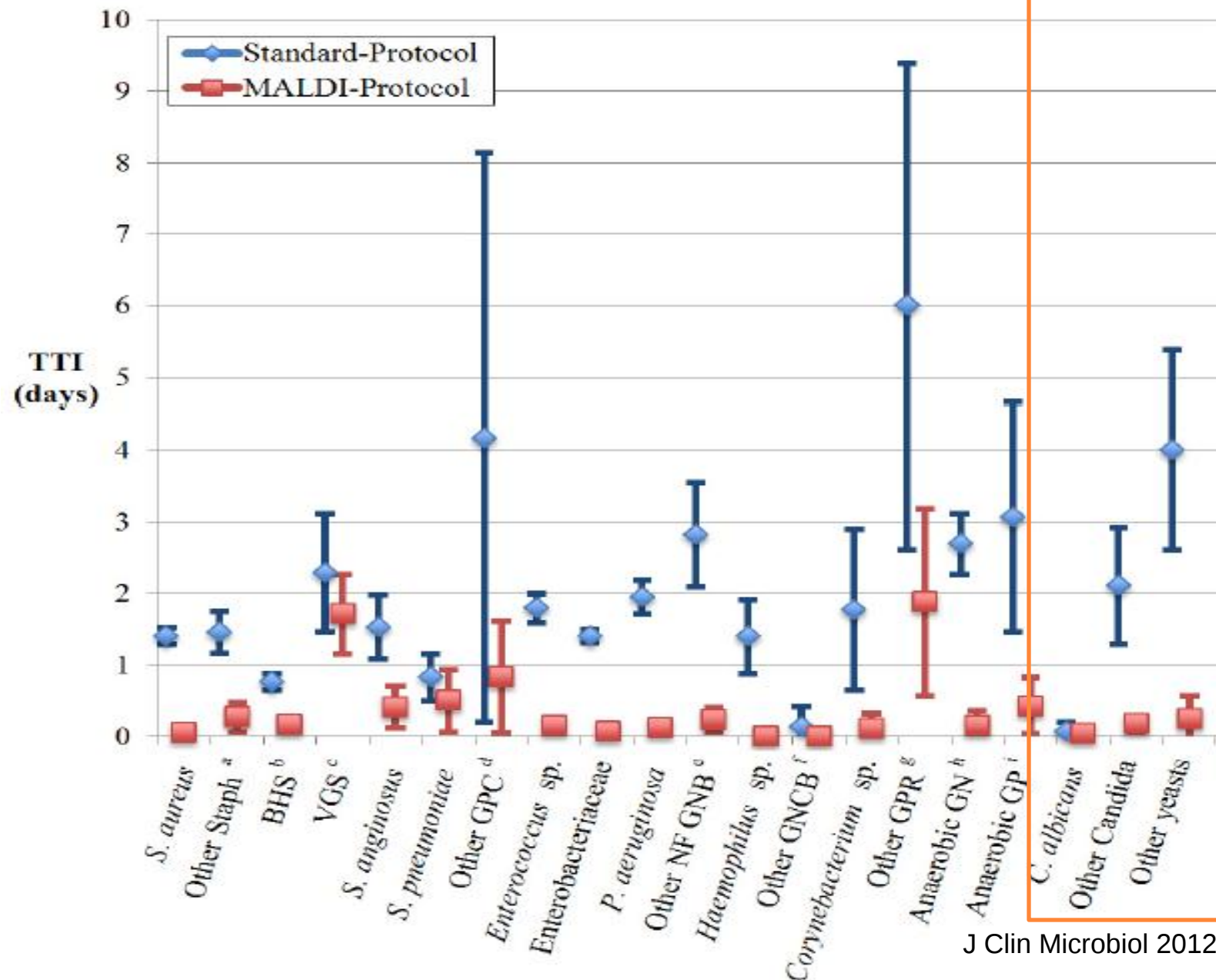
TABLE 1 Methods for analysis of isolated yeast colonies

Method	Study enrollment	Positive ID (%) ^a	Correct ID (%) ^a	Reference
Tube-based extraction (Cultured for 48 h at 30°C on SAB. One to 5 colonies were extracted using 75% EtOH, followed by suspension in a 50/50 mixture of 70% formic acid and ACN.) ^b	267 yeast isolates (250 <i>Candida</i> spp.)	247/267 (92.5)	247/247 (100)	Marklein et al. (32)
	1,192 yeast isolates (1,007 <i>Candida</i> spp.)	1,171/1,192 (98.2)	1,163/1,171 (99.3)	Bader et al. (31)
	241 yeast isolates (193 <i>Candida</i> spp.)	225/241 (93.4)	225/225 (100)	Dhiman et al. (29)
On-plate extraction (Cultured for 48 h at 30°C on SAB. One colony was transferred directly to MALDI-TOF MS analysis plate and overlaid with 70% or 25% formic acid.)	167 yeast isolates	135/167 (80.8)	135/135 (100)	Van Herendael et al. (34)
	90 yeast isolates (71 <i>Candida</i> spp.)	86/90 (95.6)	89/90 (98.9)	Theel et al. (33)
	192 yeast isolates (182 <i>Candida</i> spp.)	185/192 (96.3)	184/185 (99.5)	Iriart et al. (35)

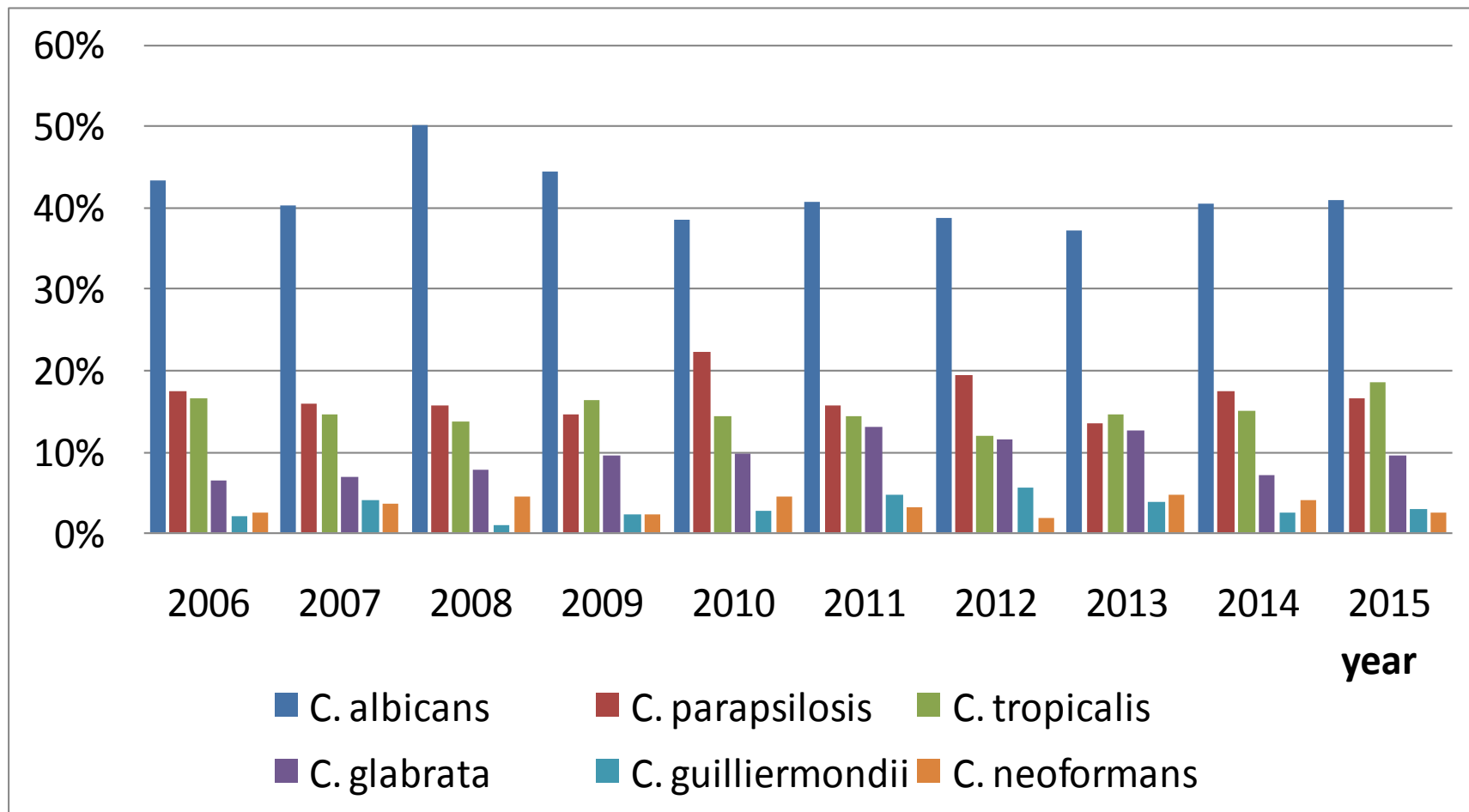
^a ID, identification. Number of isolates identified to the genus or species level (numerator) according to defined score interpretation guidelines out of the total number of isolates tested (denominator).

^b EtOH, ethanol; ACN, acetonitrile.

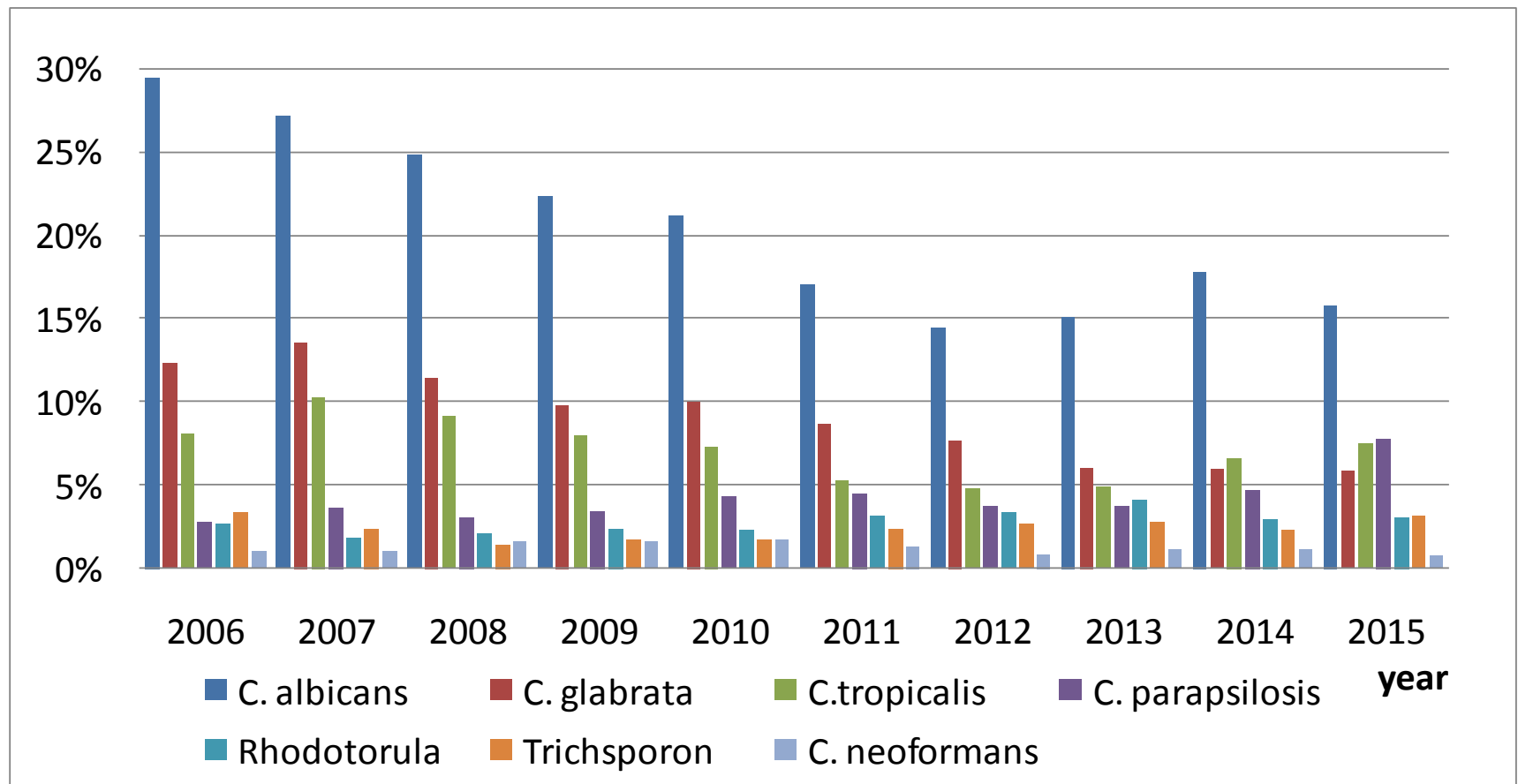
Figure 1: Time to identification (TTI) and 95% CI by MALDI- and standard-protocol



CULTURE DATA (BLOOD)



CULTURE DATA (FUNGUS)



TAXONOMY AND IDENTIFICATION

- one fungus two names
 - sexual (teleomorph) and asexual (anamorph) forms of a fungus

Anamorph name	Teleomorph name (if present)	Older name
<i>Blastoschizomyces capitatus</i>	<i>Dipodascus capitatus</i>	<i>Geotrichum capitatum</i>
<i>Candida ciferrii</i>	<i>Trichomonoascus ciferrii</i>	<i>Stephanoascus ciferrii</i>
<i>Candida famata</i>	<i>Debaryomyces hansenii</i>	
<i>Candida fermentati</i>	<i>Meyerozyma caribbica</i>	<i>Pichia caribbica</i>
<i>Candida guilliermondii</i>	<i>Meyerozyma guilliermondii</i>	<i>Pichia guilliermondii</i>
<i>Candida guilliermondii</i> var. <i>membranifaciens</i>	<i>Kodamaea ohmeri</i>	<i>Pichia ohmeri</i>
<i>Candida kefyr</i>	<i>Kluyveromyces marxianus</i>	
<i>Candida krusei</i>	<i>Pichia kudriavzevii</i>	<i>Issatchenkia orientalis</i>
<i>Candida lipolytica</i>	<i>Yarrowia lipolytica</i>	
<i>Candida lusitanae</i>	<i>Clavispora lusitanae</i>	
<i>Candida palmioleophila</i>	None	
<i>Candida pelliculosa</i>	<i>Wickerhamomyces anomalus</i>	<i>Pichia anomala</i> <i>Hansenula anomala</i>
<i>Candida utilis</i>	<i>Lindnera jadinii</i>	
<i>Cryptococcus gattii</i>	<i>Filobasidiella bacillispora</i>	<i>Cryptococcus neoformans</i> var. <i>gattii</i>
<i>Cryptococcus neoformans</i>	<i>Filobasidiella neoformans</i>	<i>Cryptococcus neoformans</i> var. <i>grubii</i> (serotype A) and <i>C. neoformans</i> var. <i>neoformans</i> (serotype D)

MALDI-TOF database

- *Candida albicans*
- *Candida albicans (africana)*
- *Candida dubliniensis*
- *Candida auris*
- *Candida glabrata*
- *Candida parapsilosis*
- *Candida tropicalis*
- *Candida krusei*
(*Issatchenkia orientalis*)
- *C. guilliermondii*
(*Meyerozyma guilliermondii*)
- *Cryptococcus gattii*
- *Cryptococcus neoformans*
- *Cryptococcus neoformans*
var_grubii
- *Cryptococcus neoformans*
var_neoformans

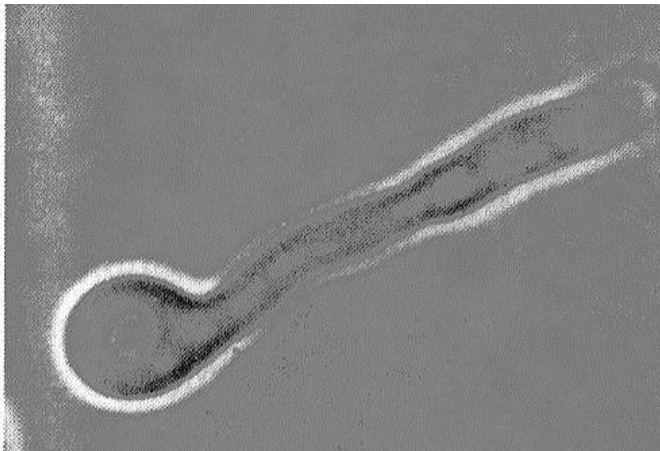
CANDIDA AND OTHER ASCOMYCETOUS YEAST



- a genus of yeasts
- the most common cause of fungal infections worldwide
- ~ 200 species
- *Candida* appears as large, round, white or cream colonies

C. albicans

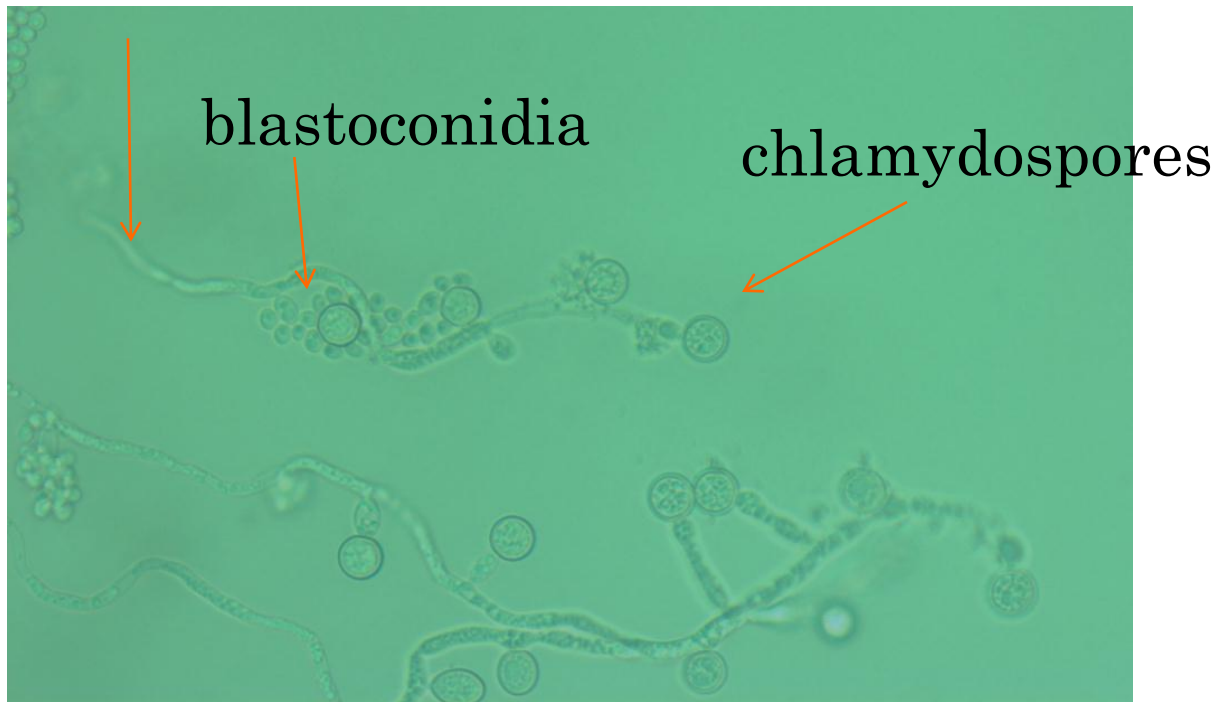
- Growth at 25, 37 and 42-45 °C
- Germ tube
- susceptible to antifungal agents



C. albicans

pseudohyphae

- On cornmeal agar
 - Blastoconidia
 - Pseudohyphae
 - Chlamydospore



C. albicans ver. africana

- susceptible to antifungal agents
- biovar of *C. albicans*
 - germ tubes (+)
 - Chlamydospores on cornmeal (-)
- Jan~Jun.2016 in CGMH

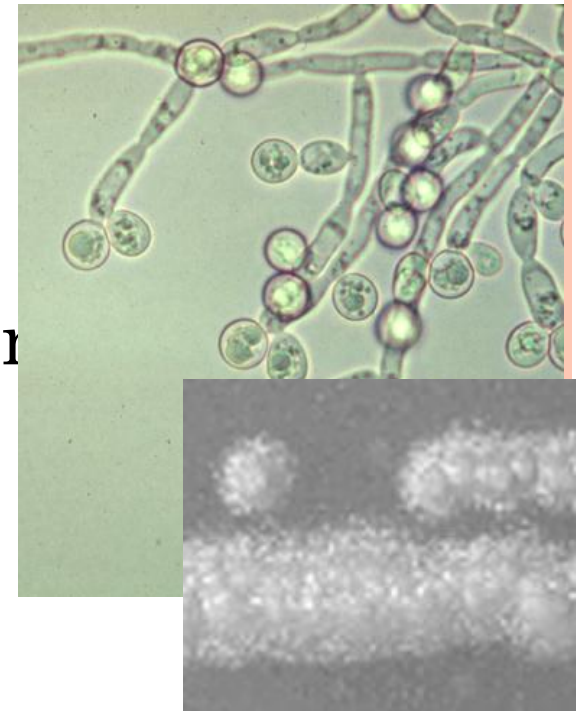
name	No.	%
<i>C. albicans</i> / <i>C. africana</i>	1011	99.5%
<i>C. dubliniensis</i>	5	0.5%

C. dubliniensis

- AIDS patient
- resistance to fluconazole (frequently or <5%)
- phenotypic similarities to *C. albicans*
 - ChromAgar
 - Germ tubes (+)
 - Chlamydospores on cornmeal (+)
- A European study - 52/2,589 isolates (2.0%).
- In USA – 15 *C. dubliniensis* / 1,251 *C. albicans* (1.2%)

C. dubliniensis

- CAP 2010-A F01(1046 participating laboratories)
 - 411 (39.4 %) → *C. dubliniensis* ✓
 - 445 (42.6 %) → *C. albicans* X



Characteristics	<i>C. albicans</i>	<i>C. dubliniensis</i>
Germ tube	+	+
Terminal Chlamydospores	Single or two	In pair, triple, cluster
Growth at 45°C	+	-
Pal's agars	Smooth colonies	hyphal fringe colonies

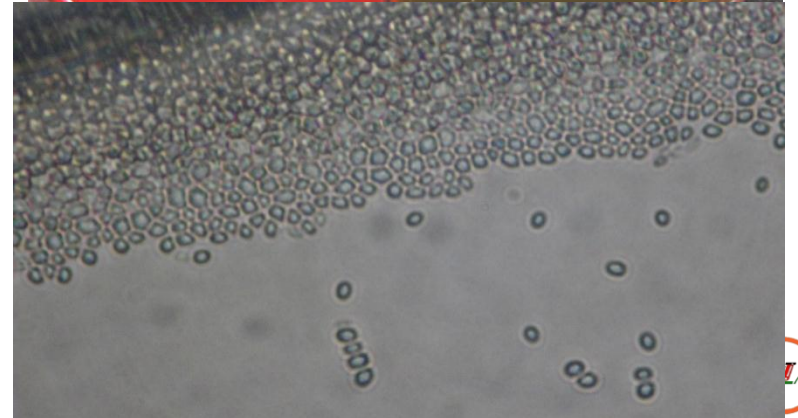
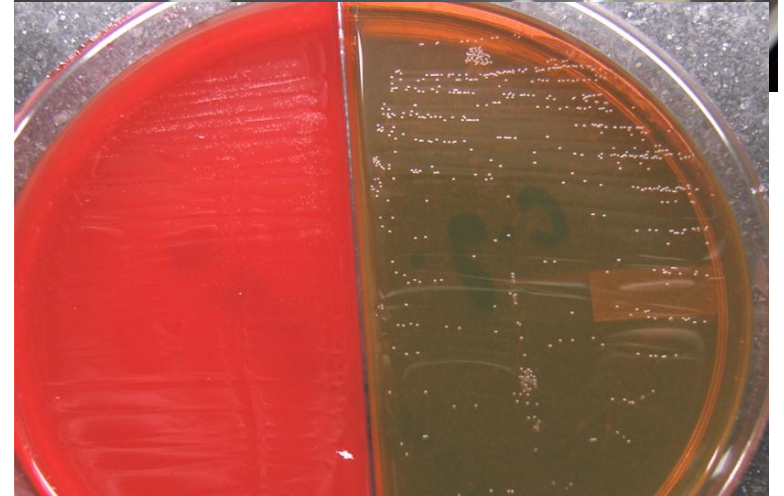
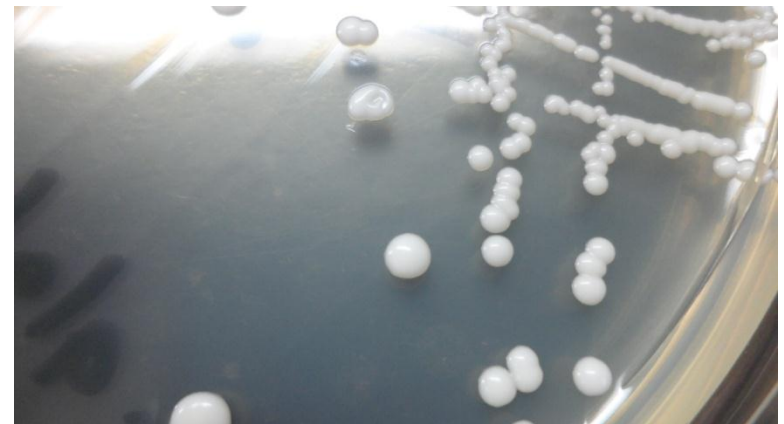
C. parapsilosis

- similar morphology (DNA-based methods)
 - *C. orthopsilosis* and *C. metapsilosis*
 - low prevalence (~1 %) in most bloodstream isolate collections surveyed
- Jan~Jun.2016 in CGMH

name	No.	%
<i>C. parapsilosis</i>	305	78.2%
<i>C. metapsilosis</i>	49	12.6%
<i>C. orthopsilosis</i>	36	9.2%

C. glabrata

- decreased susceptibility to azoles
- grows a bit more slowly than the other species of *Candida*.
- Growth better on EMB than blood
- blastospore, no pseudohyphae
- Rapid trehalose (+)



C. grabrata

- *Candida bracarensis* and *C. nivariensis* are genetically related to *C. glabrata*.
 - 1 % to 2 % of isolates
- Jan~Jun.2016 in CGMH

name	No.	%
<i>C.glabrata</i>	416	99.5%
<i>C. nivariensis</i>	2	0.5%

C. krusei
(*Issatchenkia orientalis*)

- innate resistance to fluconazole
- reduced susceptibility to amphotericin B
- pseudophyphae with elongate blastoconidia forming a cross matchsticks or treelike.



C. auris

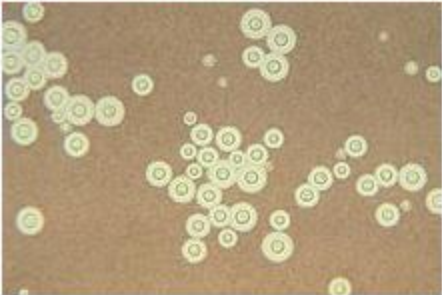
- first isolated from the external ear canal of a patient in Japan in 2009
 - bloodstream infections, wound infections, and otitis
- *Candida auris* is an emerging multidrug-resistant (MDR) yeast
 - resistant to the first-line antifungal drug fluconazole
 - rapidly evolve to develop resistance, so specific antifungal susceptibility testing is indicated
- its propensity for transmission between hospital patients.
- It has caused prolonged hospital outbreaks in India, Pakistan, Venezuela, and Colombia.

C. auris

- Germ tube (-)
- On cornmeal agar - pseudohyphae
- API AUX 20C and VITEK-2 YST -
misidentify *C. auris* as *C. haemulonii*,
Saccharomyces cerevisiae or *Rhodotorula*
glutinis.
- molecular sequencing or MALDI-TOF
- Jan~Jun.2016 in CGMH
 - No=0



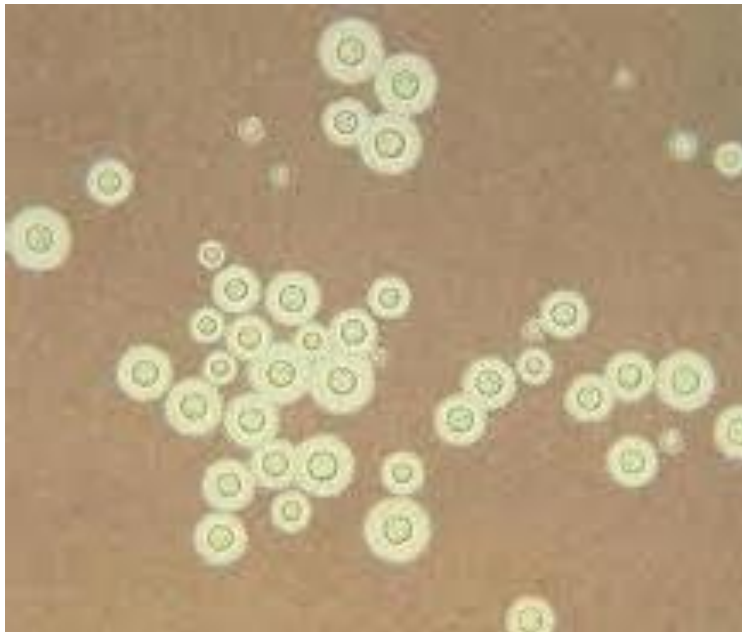
CRYPTOCOCCUS SPECIES

Cryptococcus	
	
<i>Cryptococcus neoformans</i>	
Scientific classification	
Kingdom:	Fungi
Phylum:	Basidiomycota 擔子菌門
Class:	Tremellomycetes
Order:	Tremellales
Family:	Tremellaceae
Genus:	<i>Cryptococcus</i> Vuill.
Type species	
<i>Cryptococcus neoformans</i>	
Synonyms	
<i>Filobasidiella</i>	

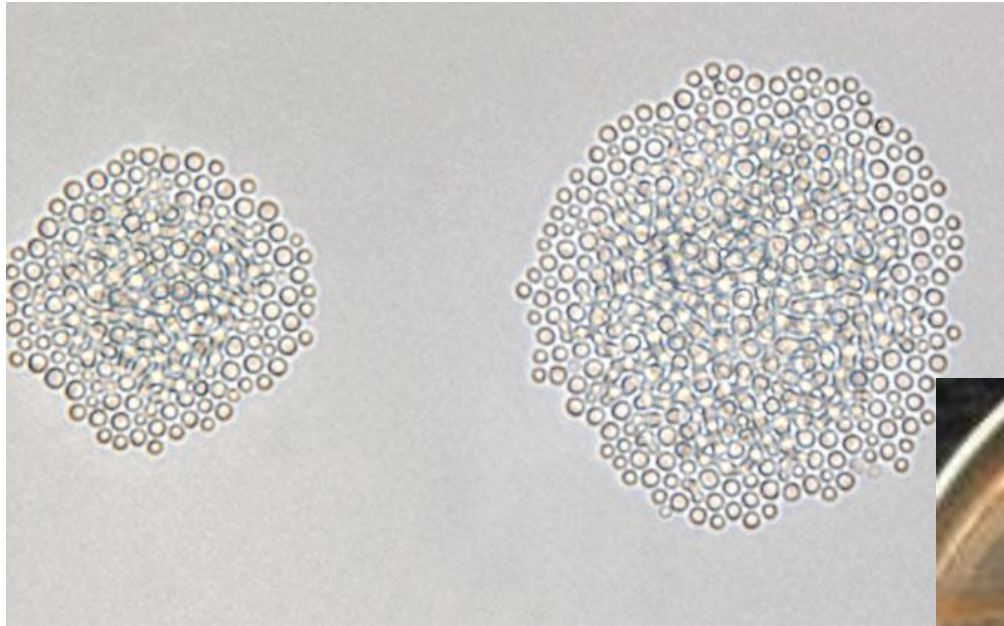
- ~37 recognized species of *Cryptococcus*
- The sexual forms or teleomorphs of *Cryptococcus* species are filamentous fungi in the genus *Filobasidiella*.
- cryptococcosis
 - *Cryptococcus neoformans*
 - *Cryptococcus gattii*

C. neoformans

- Mucoid colony
- India ink (+)



C. neoformans



No pseudohyphae

Urease(+)

phenol oxidase (+)



C. neoformans



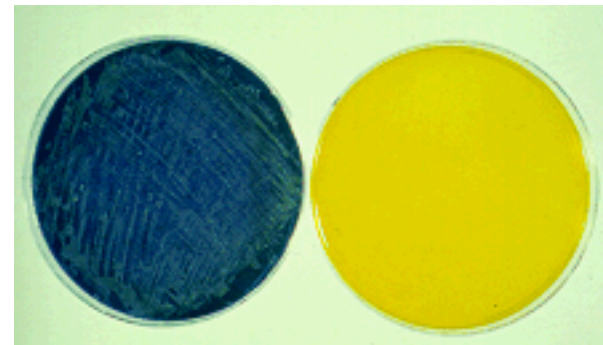
- *C. neoformans* var. *neoformans* (serotype D)
 - more geographically restricted, found mostly in Mediterranean Europe
- *C. neoformans* var. *grubii* (serotype A or hybrid serotype AD.)
 - worldwide distribution
 - in soil which has been contaminated by bird excrement
 - AIDS

name	No.	%
<i>Cryptococcus neoformans</i> <i>_var_grubii</i>	38	100%



C. gattii

- *C. gattii* (serotype B,C)
 - (formerly *Cryptococcus neoformans* var *gattii*) is endemic to tropical parts of the continent of Africa and Australia (eucalyptus)
 - causing disease (cryptococcosis) in non-immunocompromised people
- Canavanine-glycine-bromthymol blue (CGB) agar
 - Blue - *Cryptococcus gattii*



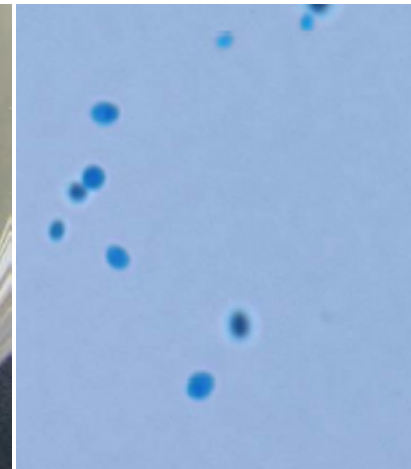
Rhodotorula

Scientific classification

Kingdom: Fungi
Division: Basidiomycota
Class: Urediniomycetes
Order: Sporidiales
Family: *incertae sedis*
Genus: ***Rhodotorula***
F.C.Harrison (1927)

Type species

Rhodotorula glutinis
(Fresen.) F.C.Harrison (1928)



- Smooth, pinkish colonies.
- Yeast morphology –forms only blastoconidia
- Urease test –will be positive.

Trichosporon

Trichosporon

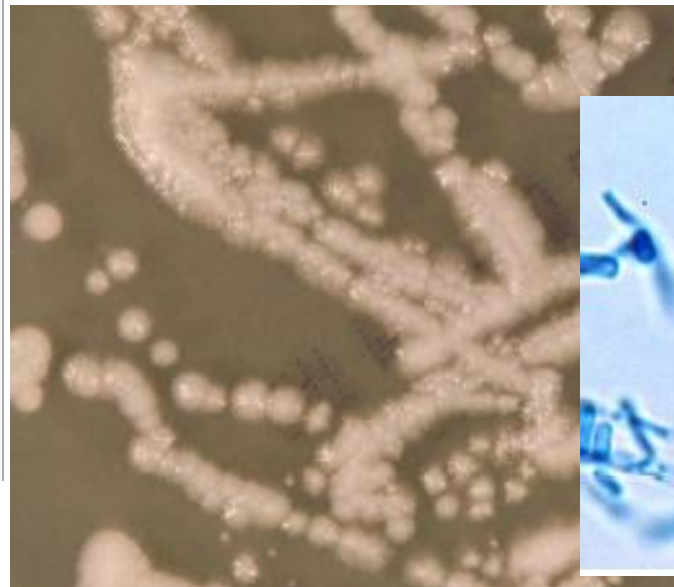
Scientific classification

Kingdom: Fungi
Phylum: Basidiomycota
Subphylum: Agaricomycotina
Class: Tremellomycetes
Order: Tremellales
Family: Trichosporonaceae
Genus: **Trichosporon**
Behrend, 1890

Type species

Trichosporon beigelia
(Küchenm. & Rabenh.) Vuill.

- Dry, rough and opaque white.
- Forms **arthroconidia**, hyphae and blastoconidia on yeast morphology
- Urease test –will be positive.

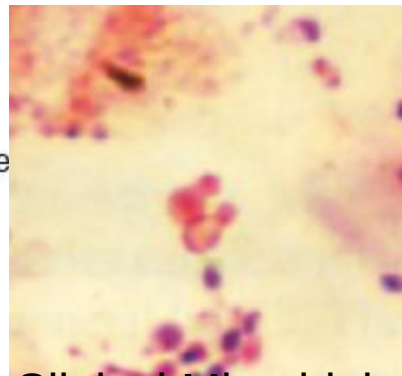
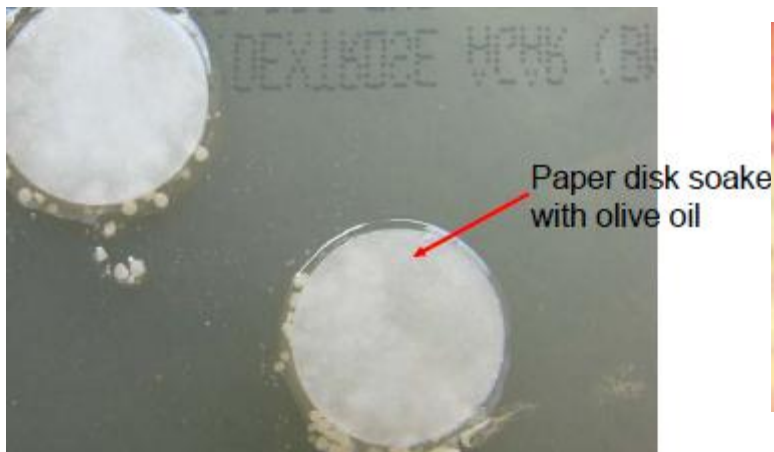


Malassezia furfur

Scientific classification

Kingdom:	Fungi
Division:	Basidiomycota
Subdivision:	Ustilaginomycotina
Class:	Exobasidiomycetes
Order:	Malasseziales
Family:	Malasseziaceae
Genus:	<i>Malassezia</i>

- Pityriasis vericolor
- needs olive oil for growth
- Slow growing
- Small white colonies
- Wet mount –shows blastoconidia with budding
- API 20C AUX - misidentified- *C. lipolytica*



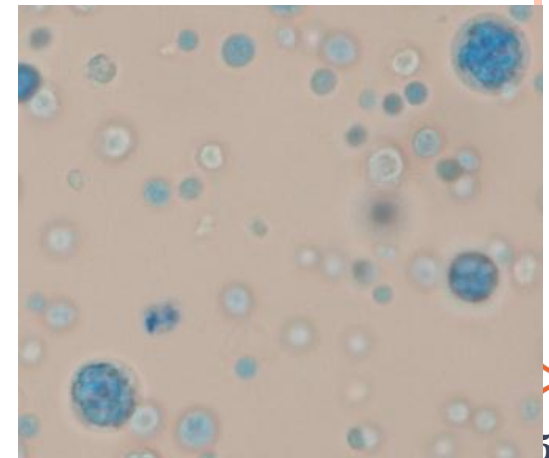
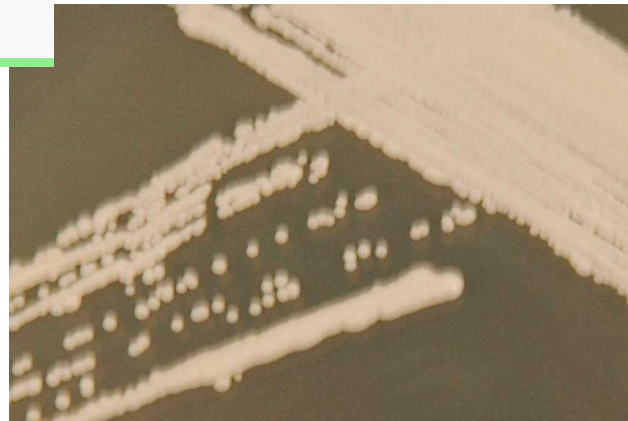
Prototheca 綠藻

Prototheca

Scientific classification

Domain: Eukaryota
Kingdom: Viridiplantae
Phylum: Chlorophyta
Class: Trebouxiophyceae
Order: Chlorellales
Family: Chlorellaceae
Genus: ***Prototheca***
Krüger, 1894^[1]

- a genus of algae
- found worldwide in sewage and soil
- Protothecosis
 - a disease found in dogs, cats, cattle, and humans
 - a wound or abrasion



JAN.2016~JUN.2016 YEAST ISOLATED FROM CGMH

name	No.	%
<i>C.albicans/C.africana</i>	1011	40.2%
<i>C. dubliniensis</i>	5	0.2%
<i>C.glabrata</i>	416	16.5%
<i>C. nivariensis</i>	2	0.1%
<i>C .tropicalis</i>	399	15.9%
<i>C. parapsilosis</i>	305	12.1%
<i>C. orthopsilosis</i>	36	1.4%
<i>C. metapsilosis</i>	49	1.9%
<i>C. guilliermondii</i>	84	3.3%
<i>C. guilliermondii</i> <i>_var_membranaefaciens</i>	3	0.1%
<i>Cryptococcus neoformans</i> <i>_var_grubii</i>	38	1.5%
<i>Rhodotorula mucilaginosa</i>	28	1.1%
<i>Trichosporon asahii</i>	25	1.0%
<i>Trichosporon mucoides</i>	21	0.8%

name	No.	%
<i>R. minuta</i>	13	0.5%
<i>Trichosporon faecale</i>	10	0.4%
<i>C. haemulonii</i>	9	0.4%
<i>C. lusitaniae</i>	8	0.3%
<i>C. krusei</i>	7	0.3%
<i>C. duobushaemulonii</i>	5	0.2%
<i>Trichosporon japonicum</i>	5	0.2%
<i>Trichosporon montevidense</i>	5	0.2%
<i>C. kefir</i>	4	0.2%
<i>C. intermedia</i>	3	0.1%
<i>C. fabianii</i>	3	0.1%
<i>C. lipolytica</i>	3	0.1%
<i>Cryptococcus diffluens</i>	3	0.1%
<i>Kluyvera ascorbata</i>	3	0.1%
other	15	0.6%
總計	2516	



THANKS FOR YOUR ATTENTION